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Lendlease Communities Pty Ltd c/- GLN Planning Level 2, 88 Phillip Street PARRAMATTA NSW 2150 Project 94571.00 26 February 2020 R.001.Rev0 EMG

Attention: Daniel Nay

Email:

Supplementary Investigation - Shed Proposed Residential Subdivision 16 Chapman Street, Werrington, NSW

1. Introduction

Douglas Partners Pty Ltd (DP) was commissioned by Lendlease Communities Pty Ltd c/- GLN Planning ('Lendlease') to undertake a supplementary contamination investigation of soils next to a shed in relation to the proposed residential subdivision of 16 Chapman Street, Werrington ('the site').

DP understands that Lendlease has submitted a development application (DA) to Council for the proposed development including previous contamination investigation reports for the site to address the requirements of State Environment Planning Policy 55 – Remediation of Land (SEPP 55). The findings of the previous reports of relevance to this investigation are provided in Section 3.3 of this report.

Council has submitted a Request for Information (RFI) to Lendlease with queries to inform the development application (DA) for the proposed residential subdivision including a request for additional assessment of impact identified by Geotesta Pty Ltd (see Section 3.3) associated with a shed and composting toilet at the site. This letter report documents the findings of the additional investigation of impact associated with the shed. It is noted that the shed and composting toilet are still located on site, therefore the additional assessment of the composting toilet will be undertaken once the structure is removed from the site.

2. Scope of Works

The scope of works carried out as part of the current investigation are as follows:

- A review of previous contamination reports was carried out (Section 3.3);
- Using hand tools, three surface soil samples were collected from next to the shed structure. The location of the soil samples collected is presented in Drawing 1, attached;



Integrated Practical Solutions

- Schedule soil samples for analysis for metals (including lead and chromium), total recoverable hydrocarbons (TRH), benzene, toluene, ethylbenzene and total xylenes (BTEX), polycyclic aromatic hydrocarbons (PAH), total phenols, organochlorine and organophosphorus pesticides (OC/OP), polychlorinated biphenyls (PCBs) and asbestos (presence/absence);
- Screen soil analytical results against human health and ecological criteria for low-density residential land use as presented in NSW EPA endorsed guidelines National Environment Protection Council (NEPC) National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended 2013 (NEPC, 2013); and
- Present the findings of the investigation in this letter report.

3. Site Background

3.1 Site Description

The site is approximately 28 hectares in size and is located in the local government area of Penrith City Council.

The site is currently unoccupied and comprises cleared bushland in the east and west and low-lying bush/mature trees in the centre and northern portion.

3.2 Soils and Geology

Reference to the Soils Landscape 1:100,000 *Penrith Sheet* indicates that the northern part of the site is located on the erosional Luddenham soil landscape and the southern part by alluvial South Creek podzolic soils/solodic soils and clays.

Reference to the Geological Series 1:100,000 *Penrith Sheet 9030* (1991) indicates that the site is underlain by Wianamatta Group Bringelly Shales of Triassic Age. This formation consists of shale, claystone, laminate and minor coal bands which are typically overlain by stiff residual clay soil.

3.3 **Previous investigations**

DP understands that Council has received several documents to address SEPP 55 including a report prepared by Geotesta Pty Ltd (Geotesta) titled *Environmental Data Gap Assessment Report for Proposed Residential Development - 16 Chapman Street, Werrington, NSW*, dated 20 October 2017, Ref: NE211 (Geotesta, 2017).

The Geotesta investigation was undertaken to further assess soil conditions below and surrounding the existing composting toilet and shed whilst the structures were still *in situ*. The scope of the investigation included a review of previous investigations, collection and analysis of nine surface samples and reporting. Of the nine samples, eight were analysed for select contaminants of potential concern (COPC).



Analytical results were assessed by Geotesta and identified elevated metals (lead and chromium) in one soil sample (BH7) which was collected from surface materials (but not soils) on the shed slab floor. Pathogens were also recorded above the laboratory limit of reporting in one sample collected by Geotesta from the toilet compost (BH9).

A comprehensive description of material sampled from BH7 was not available. Because the hardstanding shed floor was still in situ at the time of the Geotesta investigation and based on the information presented in the Geotesta report, 'surface materials' sampled by Geotesta will likely comprise residual material and dust accumulated on the shed floor, and not soils. Such surface materials are likely to be highly concentrated in metals and other contaminants when compared to soils. As such, assessment of results for BH7 against NEPC (2013) criteria is inappropriate.

As with BH7, a comprehensive description of material sampled from BH9 was not available. The sample type was described by Geotesta as 'toilet compost' which indicates that material inside the toilet was sampled by Geotesta. It is reasonable to assume that material inside the toilet includes faecal material which would also be impacted with pathogens. DP assumes that the toilet and its contents (i.e. the toilet compost) will be removed from the site prior to bulk earthworks. DP considers that the sampling and analysis of such material for a contamination investigation for the proposed development is inappropriate.

4. Results

4.1 Field Work Observations

The field work was carried out by a DP environmental scientist on 21 January 2020. The surface material sampled comprised (topsoil) brown silty clay with trace organics (leaf litter and rootlets). No odours or staining indicating potential contamination were observed at any of the sampling locations. Soil sampling locations are presented on Drawing 2.

4.2 Analytical Results

Soil analytical results have been summarised and screened against human health and ecological criteria for urban and residential land use as sourced from NEPC (2013) guidelines. The laboratory analytical reports as well as a summary table showing the results and adopted SAC are attached to this report. All soil analytical results were below the laboratory practical quantitation limit (PQL) and/or the adopted SAC.

5. Discussion and Conclusions

The scope of the current investigation included targeted sampling of surface soils next to a shed located on the site. The purpose of the investigation is to ascertain whether or not exceedances observed by Geotesta (2017) pose an actual unacceptable risk to human health and the environment with respect to the proposed residential development.



DP has undertaken a review of the Geotesta (2017) report as part of the current investigation and considers that the surface materials likely comprise dust and other residual materials that have, over time, accumulated on the shed floor. Assessment of such materials against soil criteria is inappropriate.

Soil analytical results from surface sampling undertaken next to the shed as part of the current investigation do not exceed the adopted NEPC (2013) criteria. Based on the findings of this investigation, DP considers that soil conditions surrounding the shed are suitable for the proposed residential development from a contamination perspective. As a precautionary measure, the shed and toilet including residual material present in the shed and the contents of the toilet should be removed from the site before bulk earthworks commences.

Additional sampling below the composting toilet should be undertaken at a later date once the structure has been removed from the site to confirm whether or not soils beneath the facility are impacted with pathogens.

6. Limitations

Douglas Partners Pty Ltd (DP) has prepared this report for this project at 16 Chapman Street, Werrington, NSW in accordance with DP's proposal NWS200003 dated 21 February 2020 and acceptance received from Daniel Nay of Lendlease dated 21 January 2020. The agreement for the works forms part of a larger contract currently being negotiated with Lendlease. This report is provided for the exclusive use of Lendlease for this project only and for the purposes as described in the report. It should not be used by or relied upon for other projects or purposes on the same or other site or by a third party. Any party so relying upon this report beyond its exclusive use and purpose as stated above, and without the express written consent of DP, does so entirely at its own risk and without recourse to DP for any loss or damage. In preparing this report DP has necessarily relied upon information provided by the client and/or their agents.

The results provided in the report are indicative of the sub-surface conditions on the site only at the specific sampling and/or testing locations, and then only to the depths investigated and at the time the work was carried out. Sub-surface conditions can change abruptly due to variable geological processes and also as a result of human influences. Such changes may occur after DP's field testing has been completed.

DP's advice is based upon the conditions encountered during this investigation. The accuracy of the advice provided by DP in this report may be affected by undetected variations in ground conditions across the site between and beyond the sampling and/or testing locations. The advice may also be limited by budget constraints imposed by others or by site accessibility.

This report must be read in conjunction with all of the attached and should be kept in its entirety without separation of individual pages or sections. DP cannot be held responsible for interpretations or conclusions made by others unless they are supported by an expressed statement, interpretation, outcome or conclusion stated in this report.

This report, or sections from this report, should not be used as part of a specification for a project, without review and agreement by DP. This is because this report has been written as advice and opinion rather than instructions for construction.



The contents of this report do not constitute formal design components such as are required, by the Health and Safety Legislation and Regulations, to be included in a Safety Report specifying the hazards likely to be encountered during construction and the controls required to mitigate risk. This design process requires risk assessment to be undertaken, with such assessment being dependent upon factors relating to likelihood of occurrence and consequences of damage to property and to life. This, in turn, requires project data and analysis presently beyond the knowledge and project role respectively of DP. DP may be able, however, to assist the client in carrying out a risk assessment of potential hazards contained in the Comments section of this report, as an extension to the current scope of works, if so requested, and provided that suitable additional information is made available to DP. Any such risk assessment would, however, be necessarily restricted to the environmental components set out in this report and to their application by the project designers to project design, construction, maintenance and demolition.

Please contact the undersigned if you have any questions on this matter.

Yours faithfully Douglas Partners Pty Ltd

Reviewed by

Emily McGinty Senior Associate Dean Woods Senior Associate

Attachments: About this Report Drawings 1 and 2 Laboratory Analytical Report Summary Tables



Introduction

These notes have been provided to amplify DP's report in regard to classification methods, field procedures and the comments section. Not all are necessarily relevant to all reports.

DP's reports are based on information gained from limited subsurface excavations and sampling, supplemented by knowledge of local geology and experience. For this reason, they must be regarded as interpretive rather than factual documents, limited to some extent by the scope of information on which they rely.

Copyright

This report is the property of Douglas Partners Pty Ltd. The report may only be used for the purpose for which it was commissioned and in accordance with the Conditions of Engagement for the commission supplied at the time of proposal. Unauthorised use of this report in any form whatsoever is prohibited.

Borehole and Test Pit Logs

The borehole and test pit logs presented in this report are an engineering and/or geological interpretation of the subsurface conditions, and their reliability will depend to some extent on frequency of sampling and the method of drilling or excavation. Ideally, continuous undisturbed sampling or core drilling will provide the most reliable assessment, but this is not always practicable or possible to justify on economic grounds. In any case the boreholes and test pits represent only a very small sample of the total subsurface profile.

Interpretation of the information and its application to design and construction should therefore take into account the spacing of boreholes or pits, the frequency of sampling, and the possibility of other than 'straight line' variations between the test locations.

Groundwater

Where groundwater levels are measured in boreholes there are several potential problems, namely:

 In low permeability soils groundwater may enter the hole very slowly or perhaps not at all during the time the hole is left open;

- A localised, perched water table may lead to an erroneous indication of the true water table;
- Water table levels will vary from time to time with seasons or recent weather changes. They may not be the same at the time of construction as are indicated in the report; and
- The use of water or mud as a drilling fluid will mask any groundwater inflow. Water has to be blown out of the hole and drilling mud must first be washed out of the hole if water measurements are to be made.

More reliable measurements can be made by installing standpipes which are read at intervals over several days, or perhaps weeks for low permeability soils. Piezometers, sealed in a particular stratum, may be advisable in low permeability soils or where there may be interference from a perched water table.

Reports

The report has been prepared by qualified personnel, is based on the information obtained from field and laboratory testing, and has been undertaken to current engineering standards of interpretation and analysis. Where the report has been prepared for a specific design proposal, the information and interpretation may not be relevant if the design proposal is changed. If this happens, DP will be pleased to review the report and the sufficiency of the investigation work.

Every care is taken with the report as it relates to interpretation of subsurface conditions, discussion of geotechnical and environmental aspects, and recommendations or suggestions for design and construction. However, DP cannot always anticipate or assume responsibility for:

- Unexpected variations in ground conditions. The potential for this will depend partly on borehole or pit spacing and sampling frequency;
- Changes in policy or interpretations of policy by statutory authorities; or
- The actions of contractors responding to commercial pressures.

If these occur, DP will be pleased to assist with investigations or advice to resolve the matter.

About this Report

Site Anomalies

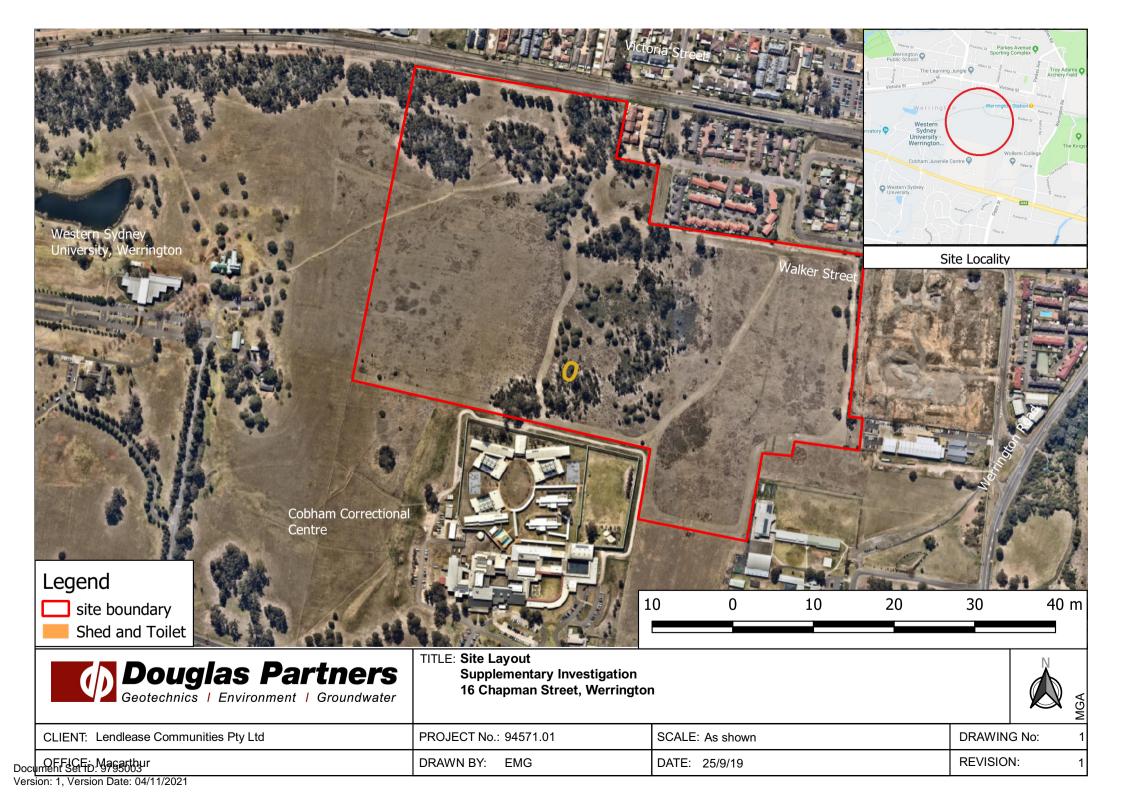
In the event that conditions encountered on site during construction appear to vary from those which were expected from the information contained in the report, DP requests that it be immediately notified. Most problems are much more readily resolved when conditions are exposed rather than at some later stage, well after the event.

Information for Contractual Purposes

Where information obtained from this report is provided for tendering purposes, it is recommended that all information, including the written report and discussion, be made available. In circumstances where the discussion or comments section is not relevant to the contractual situation, it may be appropriate to prepare a specially edited document. DP would be pleased to assist in this regard and/or to make additional report copies available for contract purposes at a nominal charge.

Site Inspection

The company will always be pleased to provide engineering inspection services for geotechnical and environmental aspects of work to which this report is related. This could range from a site visit to confirm that conditions exposed are as expected, to full time engineering presence on site.





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CERTIFICATE OF ANALYSIS 234894

| Client Details | |
|----------------|--|
| Client | Douglas Partners Pty Ltd Smeaton Grange |
| Attention | Erin Leslie, Emily McGinty |
| Address | 18 Waler Crescent, Smeaton Grange, NSW, 2567 |

| Sample Details | |
|--------------------------------------|------------------------|
| Your Reference | Council RFI Werrington |
| Number of Samples | 5 Soil |
| Date samples received | 20/01/2020 |
| Date completed instructions received | 20/01/2020 |

Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

| Report Details | | | | | | |
|---|--|--|--|--|--|--|
| Date results requested by | 28/01/2020 | | | | | |
| Date of Issue | 24/01/2020 | | | | | |
| NATA Accreditation Number 2901. This document shall not be reproduced except in full. | | | | | | |
| Accredited for compliance with IS | Accredited for compliance with ISO/IEC 17025 - Testing. Tests not covered by NATA are denoted with * | | | | | |

Results Approved By Diego Bigolin, Team Leader, Inorganics Josh Williams, Senior Chemist Loren Bardwell, Senior Chemist

Authorised By

Nancy Zhang, Laboratory Manager

Envirolab Reference: 234894 Revision No: R00

Document Set ID: 9795003 Version: 1, Version Date: 04/11/2021



| vTRH(C6-C10)/BTEXN in Soil | | | | | | |
|--|-------|------------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 | 234894-5 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 | ТВ |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil | Soil |
| Date extracted | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| TRH C ₆ - C ₉ | mg/kg | <25 | <25 | <25 | <25 | <25 |
| TRH C ₆ - C ₁₀ | mg/kg | <25 | <25 | <25 | <25 | <25 |
| vTPH C ₆ - C ₁₀ less BTEX (F1) | mg/kg | <25 | <25 | <25 | <25 | [NA] |
| Benzene | mg/kg | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 |
| Toluene | mg/kg | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| Ethylbenzene | mg/kg | <1 | <1 | <1 | <1 | <1 |
| m+p-xylene | mg/kg | <2 | <2 | <2 | <2 | <2 |
| o-Xylene | mg/kg | <1 | <1 | <1 | <1 | <1 |
| naphthalene | mg/kg | <1 | <1 | <1 | <1 | [NA] |
| Total +ve Xylenes | mg/kg | <3 | <3 | <3 | <3 | <3 |
| Surrogate aaa-Trifluorotoluene | % | 114 | 112 | 119 | 109 | 116 |

| svTRH (C10-C40) in Soil | | | | | |
|---------------------------------------|-------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil |
| Date extracted | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 23/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| TRH C ₁₀ - C ₁₄ | mg/kg | <50 | <50 | <50 | <50 |
| TRH C ₁₅ - C ₂₈ | mg/kg | <100 | <100 | <100 | <100 |
| TRH C ₂₉ - C ₃₆ | mg/kg | 180 | 150 | <100 | 210 |
| TRH >C10 -C16 | mg/kg | <50 | <50 | <50 | <50 |
| TRH >C10 - C16 less Naphthalene (F2) | mg/kg | <50 | <50 | <50 | <50 |
| TRH >C ₁₆ -C ₃₄ | mg/kg | 180 | 130 | <100 | 180 |
| TRH >C ₃₄ -C ₄₀ | mg/kg | 130 | 120 | <100 | 180 |
| Total +ve TRH (>C10-C40) | mg/kg | 320 | 250 | <50 | 350 |
| Surrogate o-Terphenyl | % | 76 | 104 | 77 | 82 |

| PAHs in Soil | | | | | |
|--------------------------------|-------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil |
| Date extracted | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Naphthalene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Acenaphthylene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Acenaphthene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Fluorene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Phenanthrene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Anthracene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Fluoranthene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Pyrene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Benzo(a)anthracene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Chrysene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Benzo(b,j+k)fluoranthene | mg/kg | <0.2 | <0.2 | <0.2 | <0.2 |
| Benzo(a)pyrene | mg/kg | <0.05 | <0.05 | <0.05 | <0.05 |
| Indeno(1,2,3-c,d)pyrene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Dibenzo(a,h)anthracene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Benzo(g,h,i)perylene | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Total +ve PAH's | mg/kg | <0.05 | <0.05 | <0.05 | <0.05 |
| Benzo(a)pyrene TEQ calc (zero) | mg/kg | <0.5 | <0.5 | <0.5 | <0.5 |
| Benzo(a)pyrene TEQ calc(half) | mg/kg | <0.5 | <0.5 | <0.5 | <0.5 |
| Benzo(a)pyrene TEQ calc(PQL) | mg/kg | <0.5 | <0.5 | <0.5 | <0.5 |
| Surrogate p-Terphenyl-d14 | % | 98 | 98 | 92 | 94 |

| Organochlorine Pesticides in soil | | | | | |
|-----------------------------------|-------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil |
| Date extracted | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| alpha-BHC | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| НСВ | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| beta-BHC | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| gamma-BHC | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Heptachlor | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| delta-BHC | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Aldrin | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Heptachlor Epoxide | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| gamma-Chlordane | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| alpha-chlordane | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Endosulfan I | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| pp-DDE | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Dieldrin | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Endrin | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Endosulfan II | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| pp-DDD | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Endrin Aldehyde | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| pp-DDT | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Endosulfan Sulphate | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Methoxychlor | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Total +ve DDT+DDD+DDE | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Surrogate TCMX | % | 119 | 117 | 110 | 110 |

| Organophosphorus Pesticides in Soil | | | | | |
|-------------------------------------|-------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil |
| Date extracted | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Dichlorvos | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Dimethoate | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Diazinon | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Chlorpyriphos-methyl | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Ronnel | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Fenitrothion | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Malathion | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Chlorpyriphos | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Parathion | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Bromophos-ethyl | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Ethion | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Azinphos-methyl (Guthion) | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Surrogate TCMX | % | 119 | 117 | 110 | 110 |

| PCBs in Soil | | | | | |
|----------------------------|-------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil |
| Date extracted | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Aroclor 1016 | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Aroclor 1221 | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Aroclor 1232 | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Aroclor 1242 | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Aroclor 1248 | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Aroclor 1254 | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Aroclor 1260 | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Total +ve PCBs (1016-1260) | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Surrogate TCMX | % | 119 | 117 | 110 | 110 |

| Acid Extractable metals in soil | | | | | |
|---------------------------------|-------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil |
| Date prepared | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Arsenic | mg/kg | 8 | 7 | 7 | 7 |
| Cadmium | mg/kg | <0.4 | <0.4 | <0.4 | <0.4 |
| Chromium | mg/kg | 23 | 21 | 19 | 19 |
| Copper | mg/kg | 13 | 12 | 9 | 12 |
| Lead | mg/kg | 30 | 25 | 18 | 22 |
| Mercury | mg/kg | <0.1 | <0.1 | <0.1 | <0.1 |
| Nickel | mg/kg | 6 | 6 | 5 | 6 |
| Zinc | mg/kg | 81 | 46 | 95 | 49 |

| Misc Soil - Inorg | | | |
|-----------------------------|-------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 |
| Your Reference | UNITS | TP1 | TP2 |
| Date Sampled | | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil |
| Date prepared | - | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 22/01/2020 | 22/01/2020 |
| Total Phenolics (as Phenol) | mg/kg | <5 | <5 |

| Moisture | | | | | |
|----------------|-------|------------|------------|------------|------------|
| Our Reference | | 234894-1 | 234894-2 | 234894-3 | 234894-4 |
| Your Reference | UNITS | TP1 | TP2 | TP3 | D1 |
| Date Sampled | | 20/01/2020 | 20/01/2020 | 20/01/2020 | 20/01/2020 |
| Type of sample | | Soil | Soil | Soil | Soil |
| Date prepared | - | 22/01/2020 | 22/01/2020 | 22/01/2020 | 22/01/2020 |
| Date analysed | - | 23/01/2020 | 23/01/2020 | 23/01/2020 | 23/01/2020 |
| Moisture | % | 20 | 20 | 12 | 27 |

| Method ID | Methodology Summary |
|-------------|---|
| AT-008 | Determination of VOCs sampled onto coconut shell charcoal sorbent tubes, that can be desorbed using carbon disulphide, and analysed by GC-MS. |
| Inorg-008 | Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours. |
| Inorg-031 | Total Phenolics by segmented flow analyser (in line distillation with colourimetric finish). Solids are extracted in a caustic media prior to analysis. |
| Metals-020 | Determination of various metals by ICP-AES. |
| Metals-021 | Determination of Mercury by Cold Vapour AAS. |
| Org-003 | Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis. |
| Org-003 | Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. |
| | F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis. |
| | Note, the Total +ve TRH PQL is reflective of the lowest individual PQL and is therefore "Total +ve TRH" is simply a sum of the positive individual TRH fractions (>C10-C40). |
| Org-006 | Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD. |
| Org-006 | Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD. Note, the Total +ve PCBs PQL is reflective of the lowest individual PQL and is therefore" Total +ve PCBs" is simply a sum of the positive individual PCBs. |
| Org-012/017 | Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. |
| Org-012/017 | Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-MS and/or GC-MS/MS. |
| | Note, the Total +ve reported DDD+DDE+DDT PQL is reflective of the lowest individual PQL and is therefore simply a sum of the positive individually report DDD+DDE+DDT. |

| Method ID | Methodology Summary |
|-------------|--|
| Org-012/017 | Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013. For soil results:- 1. 'EQ PQL'values are assuming all contributing PAHs reported as <pql actually="" and="" approach="" are="" at="" be="" calculation="" can="" conservative="" contribute="" false="" give="" given="" is="" may="" most="" not="" pahs="" positive="" pql.="" present.<br="" teq="" teqs="" that="" the="" this="" to="">2. 'EQ zero'values are assuming all contributing PAHs reported as <pql and="" approach="" are="" below="" but="" calculation="" conservative="" contribute="" false="" is="" least="" more="" negative="" pahs="" pql.<br="" present="" susceptible="" teq="" teqs="" that="" the="" this="" to="" when="" zero.="">3. 'EQ half PQL'values are assuming all contributing PAHs reported as <pql a="" above.<br="" and="" approaches="" are="" between="" conservative="" half="" hence="" least="" mid-point="" most="" pql.="" stipulated="" the="">Note, the Total +ve PAHs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PAHs" is simply a sum of the positive individual PAHs.</pql></pql></pql> |
| Org-014 | Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. |
| Org-016 | Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater. |
| Org-016 | Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater. Note, the Total +ve Xylene PQL is reflective of the lowest individual PQL and is therefore "Total +ve Xylenes" is simply a sum of the positive individual Xylenes. |

| QUALITY CONT | | Duplicate S | | | | | covery % | | | |
|--------------------------------------|-------|-------------|---------|------------|------|------|----------|------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-5 | [NT] |
| Date extracted | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Date analysed | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| TRH C ₆ - C ₉ | mg/kg | 25 | Org-016 | <25 | [NT] | | [NT] | [NT] | 93 | |
| TRH C ₆ - C ₁₀ | mg/kg | 25 | Org-016 | <25 | [NT] | | [NT] | [NT] | 93 | |
| Benzene | mg/kg | 0.2 | Org-016 | <0.2 | [NT] | | [NT] | [NT] | 88 | |
| Toluene | mg/kg | 0.5 | Org-016 | <0.5 | [NT] | | [NT] | [NT] | 88 | |
| Ethylbenzene | mg/kg | 1 | Org-016 | <1 | [NT] | | [NT] | [NT] | 95 | |
| m+p-xylene | mg/kg | 2 | Org-016 | <2 | [NT] | | [NT] | [NT] | 97 | |
| o-Xylene | mg/kg | 1 | Org-016 | <1 | [NT] | | [NT] | [NT] | 93 | |
| naphthalene | mg/kg | 1 | Org-014 | <1 | [NT] | | [NT] | [NT] | [NT] | |
| Surrogate aaa-Trifluorotoluene | % | | Org-016 | 122 | [NT] | | [NT] | [NT] | 121 | |

| QUALITY CO | QUALITY CONTROL: svTRH (C10-C40) in Soil | | | | | | | | Spike Re | covery % |
|---------------------------------------|--|-----|---------|------------|------|------|------|------|------------|----------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-5 | [NT] |
| Date extracted | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Date analysed | - | | | 23/01/2020 | [NT] | | [NT] | [NT] | 23/01/2020 | |
| TRH C ₁₀ - C ₁₄ | mg/kg | 50 | Org-003 | <50 | [NT] | | [NT] | [NT] | 80 | |
| TRH C ₁₅ - C ₂₈ | mg/kg | 100 | Org-003 | <100 | [NT] | | [NT] | [NT] | 81 | |
| TRH C ₂₉ - C ₃₆ | mg/kg | 100 | Org-003 | <100 | [NT] | | [NT] | [NT] | 88 | |
| TRH >C ₁₀ -C ₁₆ | mg/kg | 50 | Org-003 | <50 | [NT] | | [NT] | [NT] | 80 | |
| TRH >C ₁₆ -C ₃₄ | mg/kg | 100 | Org-003 | <100 | [NT] | | [NT] | [NT] | 81 | |
| TRH >C ₃₄ -C ₄₀ | mg/kg | 100 | Org-003 | <100 | [NT] | | [NT] | [NT] | 88 | |
| Surrogate o-Terphenyl | % | | Org-003 | 77 | [NT] | [NT] | [NT] | [NT] | 82 | [NT] |

| QUALI | | | Du | Duplicate | | Spike Recovery % | | | | |
|---------------------------|-------|------|-------------|------------|------|------------------|------|------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-4 | [NT] |
| Date extracted | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Date analysed | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Naphthalene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 96 | |
| Acenaphthylene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Acenaphthene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Fluorene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 98 | |
| Phenanthrene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 100 | |
| Anthracene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Fluoranthene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 98 | |
| Pyrene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 98 | |
| Benzo(a)anthracene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Chrysene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 68 | |
| Benzo(b,j+k)fluoranthene | mg/kg | 0.2 | Org-012/017 | <0.2 | [NT] | | [NT] | [NT] | [NT] | |
| Benzo(a)pyrene | mg/kg | 0.05 | Org-012/017 | <0.05 | [NT] | | [NT] | [NT] | 116 | |
| Indeno(1,2,3-c,d)pyrene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Dibenzo(a,h)anthracene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Benzo(g,h,i)perylene | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Surrogate p-Terphenyl-d14 | % | | Org-012/017 | 92 | [NT] | | [NT] | [NT] | 95 | |

| QUALITY CO | NTROL: Organo | chlorine F | Pesticides in soil | | | Du | plicate | | Spike Recovery % | | |
|---------------------|---------------|------------|--------------------|------------|------|------|---------|------|------------------|------|--|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-4 | [NT] | |
| Date extracted | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | | |
| Date analysed | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | | |
| alpha-BHC | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 104 | | |
| НСВ | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| beta-BHC | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 102 | | |
| gamma-BHC | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| Heptachlor | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 100 | | |
| delta-BHC | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| Aldrin | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 106 | | |
| Heptachlor Epoxide | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 106 | | |
| gamma-Chlordane | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| alpha-chlordane | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| Endosulfan I | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| pp-DDE | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 104 | | |
| Dieldrin | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 112 | | |
| Endrin | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 100 | | |
| Endosulfan II | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| pp-DDD | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 92 | | |
| Endrin Aldehyde | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| pp-DDT | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| Endosulfan Sulphate | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 104 | | |
| Methoxychlor | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | | |
| Surrogate TCMX | % | | Org-012/017 | 106 | [NT] | | [NT] | [NT] | 108 | | |

| QUALITY CONTRO | s Pesticides in Soil | | | Du | plicate | | Spike Recovery % | | | |
|---------------------------|----------------------|-----|-------------|------------|---------|------|------------------|------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-4 | [NT] |
| Date extracted | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Date analysed | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Dichlorvos | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 110 | |
| Dimethoate | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Diazinon | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Chlorpyriphos-methyl | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Ronnel | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 98 | |
| Fenitrothion | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 100 | |
| Malathion | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 85 | |
| Chlorpyriphos | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 104 | |
| Parathion | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 90 | |
| Bromophos-ethyl | mg/kg | 0.1 | AT-008 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Ethion | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | 98 | |
| Azinphos-methyl (Guthion) | mg/kg | 0.1 | Org-012/017 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Surrogate TCMX | % | | Org-012/017 | 106 | [NT] | | [NT] | [NT] | 108 | |

| QUALIT | | Du | plicate | | Spike Rec | overy % | | | | |
|------------------|-------|-----|---------|------------|-----------|---------|------|------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-4 | [NT] |
| Date extracted | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Date analysed | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Aroclor 1016 | mg/kg | 0.1 | Org-006 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Aroclor 1221 | mg/kg | 0.1 | Org-006 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Aroclor 1232 | mg/kg | 0.1 | Org-006 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Aroclor 1242 | mg/kg | 0.1 | Org-006 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Aroclor 1248 | mg/kg | 0.1 | Org-006 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Aroclor 1254 | mg/kg | 0.1 | Org-006 | <0.1 | [NT] | | [NT] | [NT] | 113 | |
| Aroclor 1260 | mg/kg | 0.1 | Org-006 | <0.1 | [NT] | | [NT] | [NT] | [NT] | |
| Surrogate TCMX | % | | Org-006 | 106 | [NT] | | [NT] | [NT] | 108 | |

| QUALITY CONT | | Du | | Spike Recovery % | | | | | | |
|------------------|-------|-----|------------|------------------|------|------|------|------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-5 | [NT] |
| Date prepared | - | | | 22/01/2020 | [NT] | [NT] | | [NT] | 22/01/2020 | |
| Date analysed | - | | | 22/01/2020 | [NT] | [NT] | | [NT] | 22/01/2020 | |
| Arsenic | mg/kg | 4 | Metals-020 | <4 | [NT] | [NT] | | [NT] | 103 | |
| Cadmium | mg/kg | 0.4 | Metals-020 | <0.4 | [NT] | [NT] | | [NT] | 97 | |
| Chromium | mg/kg | 1 | Metals-020 | <1 | [NT] | [NT] | | [NT] | 108 | |
| Copper | mg/kg | 1 | Metals-020 | <1 | [NT] | [NT] | | [NT] | 104 | |
| Lead | mg/kg | 1 | Metals-020 | <1 | [NT] | [NT] | | [NT] | 106 | |
| Mercury | mg/kg | 0.1 | Metals-021 | <0.1 | [NT] | [NT] | | [NT] | 90 | |
| Nickel | mg/kg | 1 | Metals-020 | <1 | [NT] | [NT] | | [NT] | 102 | |
| Zinc | mg/kg | 1 | Metals-020 | <1 | [NT] | [NT] | | [NT] | 108 | |

| QUALITY | | Du | Spike Recovery % | | | | | | | |
|-----------------------------|-------|-----|------------------|------------|------|------|------|------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-4 | [NT] |
| Date prepared | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Date analysed | - | | | 22/01/2020 | [NT] | | [NT] | [NT] | 22/01/2020 | |
| Total Phenolics (as Phenol) | mg/kg | 5 | Inorg-031 | <5 | [NT] | [NT] | [NT] | [NT] | 101 | [NT] |

| Result Definiti | ons |
|-----------------|---|
| NT | Not tested |
| NA | Test not required |
| INS | Insufficient sample for this test |
| PQL | Practical Quantitation Limit |
| < | Less than |
| > | Greater than |
| RPD | Relative Percent Difference |
| LCS | Laboratory Control Sample |
| NS | Not specified |
| NEPM | National Environmental Protection Measure |
| NR | Not Reported |

| Quality Contro | ol Definitions |
|------------------------------------|--|
| Blank | This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples. |
| Duplicate | This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable. |
| Matrix Spike | A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist. |
| LCS (Laboratory Control Sample) | This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample. |
| Surrogate Spike | Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples. |
| Australian Drinking | Water Guidelines recommend that Thermotolerant Coliform Faecal Enterococci & E Coli levels are less than |

Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.

Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

CHAIN OF CUSTODY DESPATCH SHEET

| Project No: | | | | | Suburt | | Werring | gton | | To: | Env | irolab | | | | | | | |
|-------------------------------------|-----------|------------------|-----------------------|--------------------------|------------|-------------|--------------|------------|------------|----------------------------------|-----------------------|------------------------------------|---------------------|-----------------|---------------|-----|--|--|--|
| Project Name: | | cil RFI Wer | rington | | Order I | Number | | | | 12 Ashley St, Chatswood NSW 2067 | | | | | | | | | |
| Project Manage | | | | | Sample | | Erin Le | | | Attn: | | | | | | | | | |
| Emails: | erin.le | eslie@dou | glaspartn | ers.com.au | emily.n | ncginty@ | douglas | partners | .com.au | Phone: (02) 9910 6206 | | | | | | | | | |
| Date Required: | | e day_ 🗆 | 24 hours | | ours | 72 ho | urs 🗆 | Standar | d X | Email: | | | | | | · | | | |
| Prior Storage: | 🗆 Esk | <u>ky 🗆 Frid</u> | lge 🗆 S | helved | Do sam | oles contai | n 'potentia | ' HBM? | Yes 🛛 | No 🗆 | (If YES, the | en handle, tra | l store in accordan | ce with FPM HA | JID) | | | | |
| | . e | pled | Sample Type | Container Type | | | | | Analytes | 1 | , | | | | | | | | |
| ID | Lab ID | Date Sampled | S - soil W - water | G - glass P - plastic | Suite 8 | Suite 6 | TEH/ BTEX | ¢ | | | | | | Notes/ | preservation | | | | |
| TP1 | 1 - | 20/1/20 | S | G | х | | | | | | | | | ළුකුණි ලිහිට | Marsame. | | | | |
| TP2 | 2 | .20/1/20 | S | G | Х | | | | | | | | | 1000000000 | | | | | |
| TP3 | 3 | 20/1/20 | S | G | | х | | , | E | VIROLAB | 1 | b Services Ashley St | | SUTO | | | | | |
| D1 | 4 | 20/1/20_ | S | G | | - X | | | | | Chatswool Ph: (02) | NSW 2067 9910 6200 | | | · | | | | |
| RB TB | 5 | 20/1/20 | S | G | | | | | <u>1</u> | ob No: | 2348 | 94 | | | | | | | |
| | | | | | | | | | | ate R eceiv | | 1 I I | | | | | | | |
| | | | | | | | | _ | | me Received () | | 3] | | • | | | | | |
| | | | | | | | | | Τ | emp Cool | Ambient | | | | | | | | |
| | _ | | 7 | | | | | | | ooling: 150 ecurity Int | act/Broken | None | | | | | | | |
| · · · · | | | | | | | | | | | | | | | · · · · | | | | |
| | | | | - | | | | | | | | | | | | | | | |
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| | | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | | |
| PQL (S) mg/kg | | | | | | | | | | | | ANZECO | PQLs r | req'd for all w | ater analytes | S 🗌 | | | |
| PQL = practical Metals to Analys | | | | | t to Labo | ratory Me | thod Dete | ection Lim | nit | Lab R | eport/Re | ference N | o: | | | | | | |
| Total number of | | | | | quished | by: Erir | Leslie | Transpo | rted to la | borator | vbv: | | | | | | | | |
| Send Results to | in po | ouglas Rarti | | d Addr | ess 8 Wa | aler Cres | , Smeator | Grange | NSW | | | Phone: | | Fax: | | | | | |
| Signed: | < < | Sle. | | Received b | | | 2 | | | | Date & 7 | | 0/12 | | | | | | |

Douglas Partners Geotechnics | Environment | Groundwater

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Envirolab Services Pty Ltd ABN 37 112 535 645 12 Ashley St Chatswood NSW 2067 ph 02 9910 6200 fax 02 9910 6201 customerservice@envirolab.com.au www.envirolab.com.au

SAMPLE RECEIPT ADVICE

| Client Details | |
|----------------|---|
| Client | Douglas Partners Pty Ltd Smeaton Grange |
| Attention | Erin Leslie, Emily McGinty |

| Sample Login Details | |
|--------------------------------------|------------------------|
| Your reference | Council RFI Werrington |
| Envirolab Reference | 234894 |
| Date Sample Received | 20/01/2020 |
| Date Instructions Received | 20/01/2020 |
| Date Results Expected to be Reported | 28/01/2020 |

| Sample Condition | |
|--|----------|
| Samples received in appropriate condition for analysis | Yes |
| No. of Samples Provided | 5 Soil |
| Turnaround Time Requested | Standard |
| Temperature on Receipt (°C) | 19.2 |
| Cooling Method | Ice Pack |
| Sampling Date Provided | YES |

Comments Nil

Please direct any queries to:

| Aileen Hie | Jacinta Hurst |
|------------------------------|--------------------------------|
| Phone: 02 9910 6200 | Phone: 02 9910 6200 |
| Fax: 02 9910 6201 | Fax: 02 9910 6201 |
| Email: ahie@envirolab.com.au | Email: jhurst@envirolab.com.au |

Analysis Underway, details on the following page:



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| Sample ID | VTRH(C6-C10)/BTEXN in Soil | svTRH (C10-C40) in Soil | PAHs in Soil | Organochlorine Pesticides in soil | Organophosphorus Pesticides in Soil | PCBsin Soil | Acid Extractable metalsin soil | Misc Soil - Inorg | Asbestos ID - soils |
|-----------|----------------------------|-------------------------|--------------|-----------------------------------|--|--------------|--------------------------------|-------------------|---------------------|
| TP1 | \checkmark | \checkmark | \checkmark | \checkmark | ✓ | \checkmark | \checkmark | \checkmark | \checkmark |
| TP2 | \checkmark | \checkmark | \checkmark | \checkmark | \checkmark | \checkmark | ✓ | \checkmark | \checkmark |
| TP3 | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | | ✓ |
| D1 | \checkmark | \checkmark | \checkmark | \checkmark | \checkmark | \checkmark | ✓ | | \checkmark |
| ТВ | \checkmark | | | | | | | | |

The ' \checkmark ' indicates the testing you have requested. THIS IS NOT A REPORT OF THE RESULTS.

Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.



Table 1: Summary of Laboratory Results – Metals, TRH, BTEX, PAH

| | | | Metals | | | | | | | | | | | TRH | | | | | | | РАН | | | |
|-----------|---------------|--------------|---------|---------|----------------|----------|----------|------------------------|----------|----------|--------------|--------------|------------------------|------------------------------------|---------------|---------------|---------|---------|--------------|---------------|--------------------------|-------------------------|-----------------------|------------|
| | | | Arsenic | Cadmium | Total Chromium | Copper | Lead | Mercury (inorganic) | Nickel | Zinc | TRH C6 - C10 | TRH >C10-C16 | F1 ((С6-С10)- ВТЕХ) | F2 (>C10-C16 less Naphthalene) | F3 (>C16-C34) | F4 (>C34-C40) | Benzene | Toluene | Ethylbenzene | Total Xylenes | Naphthalene ^b | Benzo(a)pyrene (BaP) | Benzo(a)pyrene TEQ | Total PAHs |
| | | PQL | 4 | 0.4 | 1 | 1 | 1 | 0.1 | 1 | 1 | 25 | 50 | 25 | 50 | 100 | 100 | 0.2 | 0.5 | 1 | 1 | 1 | 0.05 | 0.5 | 0.05 |
| Sample ID | Depth | Sampled Date | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg |
| TP1 | 0m | 20/01/2020 | 8 | <0.4 | 23 | 13 | 30 | <0.1 | 6 | 81 | <25 | <50 | <25 | <50 | 180 | 130 | <0.2 | <0.5 | <1 | <1 | <1 | <0.05 | <0.5 | <0.05 |
| | | 20,01,2020 | 100 100 | 20 NC | 100 640 | 6000 55 | 300 1100 | 40 NC | 400 35 | 7400 150 | NC NC | NC NC | 50 180 | 280 120 | NC 1300 | NC 5600 | 0.7 65 | 480 105 | NL 125 | 110 45 | 5 170 | NC 0.7 | 3 NC | 300 NC |
| TP2 | 0m | 20/01/2020 | 7 | <0.4 | 21 | 12 | 25 | <0.1 | 6 | 46 | <25 | <50 | <25 | <50 | 130 | 120 | <0.2 | <0.5 | <1 | <1 | <1 | <0.05 | <0.5 | <0.05 |
| | UIII | 20/01/2020 | 100 100 | 20 NC | 100 640 | 6000 55 | 300 1100 | 40 NC | 400 35 | 7400 150 | NC NC | NC NC | 50 180 | 280 120 | NC 1300 | NC 5600 | 0.7 65 | 480 105 | NL 125 | 110 45 | 5 170 | NC 0.7 | 3 NC | 300 NC |
| D1 | 0m | 20/01/2020 | 7 | <0.4 | 19 | 12 | 22 | <0.1 | 6 | 49 | <25 | <50 | <25 | <50 | 180 | 180 | <0.2 | <0.5 | <1 | <1 | <1 | <0.05 | <0.5 | <0.05 |
| | UII | 20/01/2020 | 100 100 | 20 NC | 100 640 | 6000 55 | 300 1100 | 40 NC | 400 35 | 7400 150 | NC NC | NC NC | 50 180 | 280 120 | NC 1300 | NC 5600 | 0.7 65 | 480 105 | NL 125 | 110 45 | 5 170 | NC 0.7 | 3 NC | 300 NC |
| TP3 | 0m | 20/01/2020 | 7 | <0.4 | 19 | 9 | 18 | <0.1 | 5 | 95 | <25 | <50 | <25 | <50 | <100 | <100 | <0.2 | <0.5 | <1 | <1 | <1 | <0.05 | <0.5 | <0.05 |
| | 0m 20/01/2020 | 100 100 | 20 NC | 100 640 | 6000 55 | 300 1100 | 40 NC | 400 35 | 7400 150 | NC NC | NC NC | 50 180 | 280 120 | NC 1300 | NC 5600 | 0.7 65 | 480 105 | NL 125 | 110 45 | 5 170 | NC 0.7 | 3 NC | 300 NC | |

Lab result
HIL/HSL value EIL/ESL value

HIL/HSL exceedance 📕 EIL/ESL exceedance 📕 HIL/HSL and EIL/ESL exceedance 📕 ML exceedance 📕 ML and HIL/HSL or EIL/ESL exceedance

Indicates that asbestos has been detected by the lab below the PQL, refer to the lab report **Blue** = DC exceedance

Bold = Lab detections NT = Not tested NL = Non limiting NC = No criteria NA = Not applicable NAD = No asbestos detected

Notes:

| HIL/HSL/DC | NEPC, Schedule B1 - HIL A (Residential / Low - High Density), HSL A/B (Residential / Low - High Density), DC HSL A (Direct contact HSL A Residential (Low density)) |
|------------|---|
| EIL/ESL | NEPC, Schedule B1 - EIL UR/POS (Urban Residential and Public Open Space), ESL UR/POS (Urban Residential and Public Open Space) |
| ML | NEPC, Schedule B1 - ML R/P/POS (Residential, Parkland and Public Open Space) |
| а | QA/QC replicate of sample listed directly below the primary sample |
| b | reported naphthalene laboratory result obtained from BTEXN suite |
| С | criteria applies to DDT only |

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Table 2: Summary of Laboratory Results – Phenol, OCP, OPP, PCB

| | | | Phenol | | | | | | OCP | | | | | | OPP | PCB |
|-----------|-------|--------------|--------------|------------------|---------------|---------------|----------------|-------------------|-----------------|------------------|---------------|--------------|---------------|----------------|----------------|--------------|
| | | | Phenol | DDT+DDE+DDD c | DDD | DDE | DDT | Aldrin & Dieldrin | Total Chlordane | Total Endosulfan | Endrin | Heptachlor | НСВ | Methoxychlor | Chlorpyriphos | Total PCB |
| | | PQL | 5 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 |
| Sample ID | Depth | Sampled Date | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg | mg/kg |
| TP1 | 0m | 20/01/2020 | <5 100 NC | <0.1 240 180 | <0.1 NC NC | <0.1 NC NC | <0.1 NC 180 | <0.1 6 NC | <0.1 50 NC | <0.1 270 NC | <0.1 10 NC | <0.1 6 NC | <0.1 10 NC | <0.1 300 NC | <0.1 160 NC | <0.1 1 NC |
| TP2 | 0m | 20/01/2020 | <5 100 NC | <0.1 240 180 | <0.1 NC NC | <0.1 NC NC | <0.1 NC 180 | <0.1 6 NC | <0.1 50 NC | <0.1 270 NC | <0.1 10 NC | <0.1 6 NC | <0.1 10 NC | <0.1 300 NC | <0.1 160 NC | <0.1 1 NC |
| D1 | 0m | 20/01/2020 | NT 100 NC | <0.1 240 180 | <0.1 NC NC | <0.1 NC NC | <0.1 NC 180 | <0.1 6 NC | <0.1 50 NC | <0.1 270 NC | <0.1 10 NC | <0.1 6 NC | <0.1 10 NC | <0.1 300 NC | <0.1 160 NC | <0.1 1 NC |
| TP3 | 0m | 20/01/2020 | NT 100 NC | <0.1 240 180 | <0.1 NC NC | <0.1 NC NC | <0.1 NC 180 | <0.1 6 NC | <0.1 50 NC | <0.1 270 NC | <0.1 10 NC | <0.1 6 NC | <0.1 10 NC | <0.1 300 NC | <0.1 160 NC | <0.1 1 NC |

Lab result
HIL/HSL value EIL/ESL value

📙 HIL/HSL exceedance 📕 EIL/ESL exceedance 📕 HIL/HSL and EIL/ESL exceedance 📕 ML exceedance 📕 ML and HIL/HSL or EIL/ESL exceedance

Indicates that asbestos has been detected by the lab below the PQL, refer to the lab report **Blue** = DC exceedance

Bold = Lab detections NT = Not tested NL = Non limiting NC = No criteria NA = Not applicable NAD = No asbestos detected

Notes:

HIL/HSL/DC NEPC, Schedule B1 - HIL A (Residential / Low - High Density), HSL A/B (Residential / Low - High Density), DC HSL A (Direct contact HSL A Residential (Low density))

EIL/ESL NEPC, Schedule B1 - EIL UR/POS (Urban Residential and Public Open Space), ESL UR/POS (Urban Residential and Public Open Space)

- ML NEPC, Schedule B1 ML R/P/POS (Residential, Parkland and Public Open Space)
- a QA/QC replicate of sample listed directly below the primary sample
- b reported naphthalene laboratory result obtained from BTEXN suite
- c criteria applies to DDT only